Memo

DILLON

To:	In accordance with s.40 of ATIPPA
From:	Claire Smith, Dillon Consulting Limited
cc:	Matt Gosse, Dillon Consulting Limited
Date:	July 17, 2026
Subject:	101A Nearys Pond Road Waterbody and Indeterminate Stream Assessment
Our File:	26-3638

Introduction

Dillon Consulting Limited (Dillon) was retained by the Town of Portugal Cove-St. Philip's to resolve a regulatory boundary uncertainty at No. 101A Nearys Pond Road, resulting from unverified 2022 desktop hydrology mapping. A pond on the eastern edge of the property and an associated indeterminate stream have been identified as waterbodies on the Town's Environmental Protection Overlap Map (Schedule E). The definition of a waterbody in the Town's Municipal Plan and Development Regulations 2014-2024 follows the Government of Newfoundland and Labrador's definition in the Water Resources Act:

> *"A surface or subterranean source of fresh or salt water within the jurisdiction of the province, whether that source usually contains liquid or frozen water or not, and includes water above the bed of the sea that is within the jurisdiction of the province, a river, stream, brook, creek, watercourse, lake, pond, spring, lagoon, ravine, gully, canal, wetland and other flowing or standing water and the land occupied by that body of water".*

67382 Newfoundland and Labrador Limited (the property owner) has identified the waterbody and stream as artifacts of a historical poultry operation rather than naturally occurring, regulated features. To address the property owner's assertion that the pond is an artificial feature and should not be identified as a waterbody, Dillon completed an independent, evidence-based habitat assessment of the waterbody and its associated stream. This assessment provides a description of the biophysical properties of both features through field-based surveys with georeferenced photographs; environmental DNA (eDNA) sampling to assess fish presence; and information provided through previous ESAs and the historical information provided by the property owner.

Historical Information

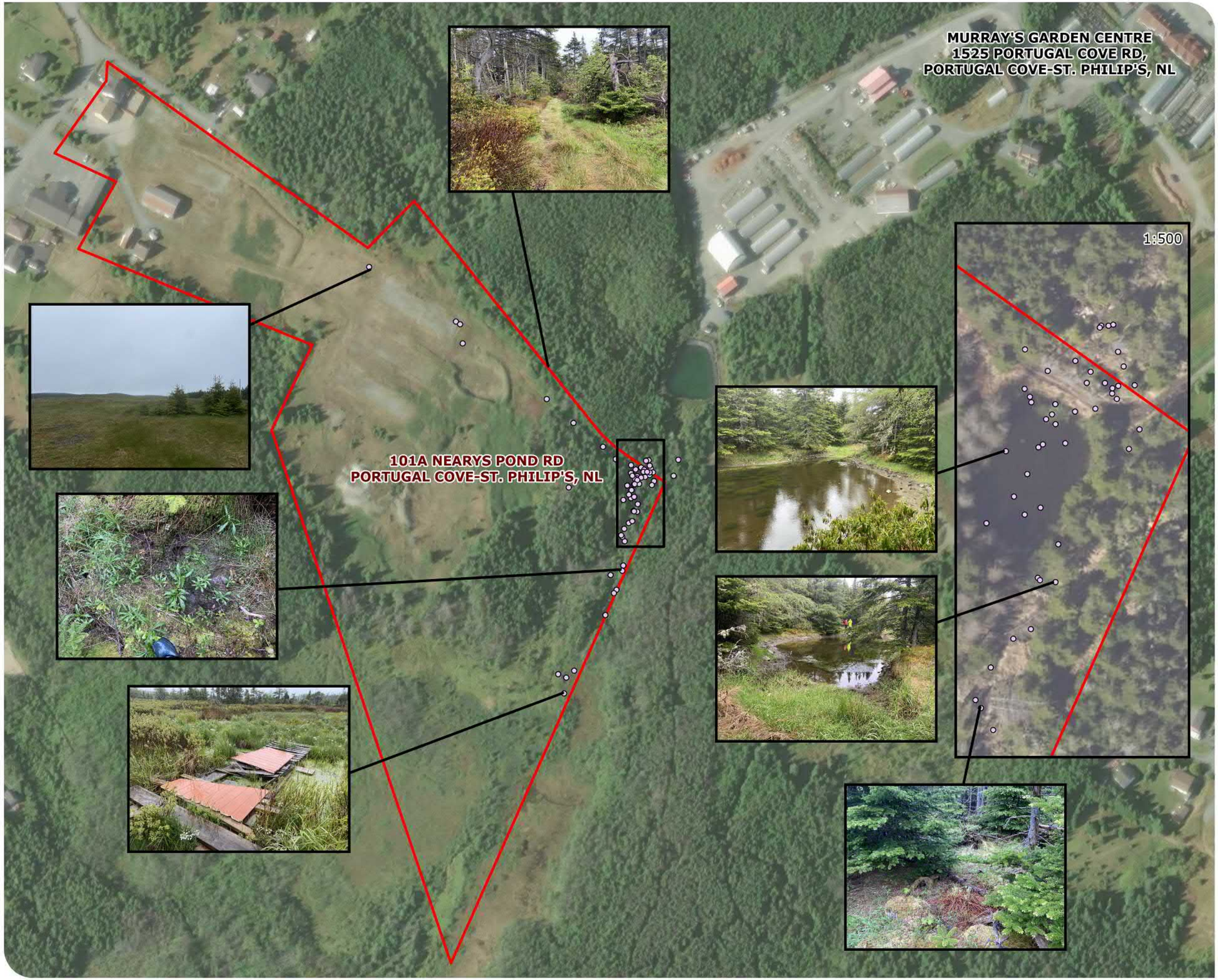
The property was previously owned by Munn Poultry Farms, a poultry egg farm. In 2024, Englobe Corp. was retained by the property owner to perform Phase I and II Environmental Site Assessments (ESAs) for due diligence purposes. It was observed that the pond on the eastern edge of the property appeared hydrologically connected to the retention pond on the nearby, upgradient property to the northeast owned by Murray's Garden Centre (Figure 1). Groundwater was inferred to flow south through the property, with much of the site sloping toward a wetland, located approximately 100m downstream, at the southern edge and a small portion sloping to the pond or to a drainage channel running through the site. The Phase II ESA was performed in response to concerns about discharge from the ponds, associated with historical farming on the site and current operations on the northeast property.

To assess potential environmental impacts to water and sediment, one water sample was collected at the southern end of the pond, near the identified outflow, and one sediment sample was collected from the top 0.10 m of sediment from shoreline at the pond's approximate centre. The surface water sample exceeded the Newfoundland and Labrador Environmental Control Water and Sewer Regulations limit for Total Suspended Solids (TSS), but all other concentrations were below regulatory limits. The Phase II ESA concluded that any impacts on surface water and sediment from current and/or historical operations on the site and up-gradient properties were negligible at the time of the study. Conclusions were based on the site being used for residential purposes with a non-potable water source.

2026 Field Methodologies

Habitat Assessment

The Dillon field team performed a preliminary site visit on June 8, 2026, along with the property owner to assess features. On June 17, 2026, the field team returned to survey the mapped waterbody and stream. The team assessed the features' physical characteristics, hydrological connectivity, and ecological function. This included collecting eDNA samples to non-invasively assess potential fish presence in the pond. The team walked north and south of the pond along the mapped indeterminate stream to inspect for active flows, connectivity to Nearys Pond or adjacent networks, and structural signs of modification. Using georeferenced photographs, field staff documented pond substrate composition and depth, potential inflows and outflows, any debris or evidence of human activity near the pond, and any signs of wildlife utilizing the immediate area.



WATERBODY AND INDETERMINATE STREAM

TOWN OF PORTUGAL COVE
ST. PHILIPS

FIELD SURVEY RESULTS

FIGURE 1

- Photo Observation Point
- Approximate Property Boundary



SCALE 1:2,500

0 25 50 100 Meters

N
W E
S

MAP DRAWING INFORMATION:
DATA PROVIDED BY DILLON CONSULTING LIMITED AND ESRI

MAP CREATED BY: HF
MAP CHECKED BY: MG
MAP PROJECTION: NAD 1983 UTM Zone 22N

DILLON

PROJECT: 26-3638
STATUS: DRAFT
DATE: 2026-07-16

FILE LOCATION: K:\2026\263638\Product\Client\PortugalFigures\PortugalFigures.aprx; LAYOUT: P263638_Fig1_FieldSurveyResults_20260716

eDNA Analysis

Environmental DNA (eDNA) analysis is a biodiversity monitoring tool that can detect organisms non-invasively using the DNA they shed into the environment, through skin cells, hair, scales, waste, and more. DNA from environmental samples, including water, sediment or soil, can be extracted, amplified and analyzed by an accredited laboratory to detect organisms of interest. Dillon conducted eDNA sampling to test for the presence of Brook Trout (*Salvelinus fontinalis*) and American Eel (*Anguilla rostrata*) in the identified waterbody. These species were chosen for their prevalence in freshwater streams and ponds in the area.

Three 1-L surface water samples were collected along the perimeter of the pond on June 17, 2026. Since eDNA distribution can be spotty, samples were collected as composites from at least three adjacent areas. Samples were collected by trained personnel into lab-supplied plastic bottles, stored in a cooler on ice, and shipped the same day to Mérieux NutriSciences for filtration and analysis. Mérieux NutriSciences, located in Guelph, Ontario, is accredited by the Standards Council of Canada (SCC) to ISO/IEC 17025 for eDNA testing.

Samples underwent DNA extraction and qPCR analysis to test for the presence of eDNA from the two target species. For each sample, eight technical replicates were used for the target species assay. A quality control assay (IntegritE-DNA) was performed using four technical replicates from each sample to assess the quality of eDNA in samples and the likelihood that target eDNA will be detected, if present. Laboratory negative and positive controls were included throughout DNA extraction and qPCR analysis to test for potential sample contamination during handling and to assess qPCR performance.

2026 Field Results

Habitat

Survey of Identified Waterbody

At the time of the survey on June 17, 2026, the waterbody consisted of clear, shallow water approximately 1 m deep. The waterbody's substrate was composed of fine sediment, mud, tufts of algae, and organic matter. The shoreline consisted of mud and vegetation, occasional cobbles and small boulders, and contained relatively little aquatic vegetation. Two water lily (*Nuphar variegata*) plants were growing near the northwest perimeter of the waterbody. The overall shape of the waterbody was roughly rectangular, with steep banks along the eastern perimeter, consistent with having been dug out by machinery rather than a natural formation. The pond appeared to have been shored up along the sides, with treed berms observed around the perimeter. A trench was carved out of the berm at the northeast perimeter of the pond, covered by two wood boards. Several large boulders and an object made of concrete and wood were observed near the northeast perimeter. A pipe ran from the pond

into the northwest shoreline, in the direction of a structure identified by the property owner as an old pump house. Inside the pump house, a section of pipe ran straight up from the floor. A potential cap for water intake was observed lying on the substrate in the northeastern quadrant of the pond, near the pipe.

Site Photographs are found in **Attachment A**.

Survey of Identified Indeterminate Stream

The team followed areas of likely water flow both north (upstream) and south (downstream) of the waterbody. Neither area contained clear stream banks indicative of consistent waterflow. There was also no surface water observed at the time of survey in any reach of the indeterminate stream that was assessed. Both areas contained areas of hydrophilic vegetation such as *Sphagnum* moss, grasses, and sedges. The distinction between conifer forest and clearings with hydrophilic vegetation was stronger in the area south of the pond, which also contained occasional wet, muddy patches and after 100 m opened into a wetland at the southern perimeter of the pond. Any hydrological connection between the excavated pond and the downstream wetland is likely limited to intermittent overland sheet flow during extreme peak-flow events or following spring snowmelt.

Fish and Wildlife Observations

No fish were observed in the pond. Tadpoles were observed in the pond during the site visit on June 8, 2026. On June 17, 2026, no tadpoles or frogs were observed but moose droppings and tracks were seen while surveying the pond's potential outlet to the south.

eDNA

Water sampling for eDNA analysis failed to detect the presence of Brook Trout or American Eel within the waterbody. Neither species was positively detected in any of the 8 technical replicates run from the three samples taken. All samples passed laboratory criteria for the IntegritE-DNA assay, indicating that any eDNA from target taxa would likely be of sufficient quality to be detected. All laboratory negative and positive controls passed laboratory criteria, indicating that no contamination was introduced during sample handling and that any negative results were not related to poor qPCR performance, respectively.

Certificates of analysis are presented in **Attachment B**.

Discussion and Assessment

A field survey of the pond's appearance noted several characteristics consistent with the pond having been constructed for agriculture operations, including a generally rectangular shape, berms around its perimeter, the placement of large boulders and human-made objects around the waterbody, and sheer, straight-edged banks that appear carved out rather than naturally formed. Additionally, the presence of a hose running from the pond toward the north, as well as a submerged pipe leading to a structure identified as a pump house, suggest the pond has been used as a water source in the past.

The pond appears to have minimal connectivity to other water bodies. The potential inlet and outlet areas surveyed north and south of the pond contained hydrophilic vegetation as well as damp areas in the potential outlet that are consistent with historic or occasional water flow. However, both areas lacked clear stream beds that indicate consistent water flow and surface water was not observed.

Additionally, the pond is unlikely to support fish. The eDNA analysis failed to detect eDNA from either Brook Trout or American Eel, two species which are common in waterbodies on the Avalon Peninsula. While eDNA sampling cannot confirm the absence of these species, the pond's shallow waters and potential for drying up seasonally ([redacted]) make it unlikely to be suitable fish habitat.

In accordance with s.40 of ATIPPA

From a field-based review of the pond's biophysical features, including its rectangular appearance with unnaturally sheer banks and high berms, its minimal connectivity to other water bodies, and the likely absence of fish, the identified waterbody and its associated stream do not appear to be natural waterbodies and likely have limited ecological function.

Closure

This report was prepared by Dillon Consulting Limited (Dillon) on behalf of 67382 NL and Labrador Limited. Dillon has used the degree of care and skill ordinarily exercised under similar circumstances at the time the work was performed by reputable members of the environmental consulting profession practicing in Canada. Dillon assumes no responsibility for conditions which were beyond its scope of work. There is no warranty expressed or implied by Dillon.

The material in the report reflects Dillon's best judgment considering the information available to Dillon at the time of preparation. Any use which a third party makes of this report, or any reliance on or decisions made based on it, are the responsibilities of such third parties. Dillon accepts no responsibility for damages, if any, suffered by any third party because of decisions made or actions based on this report.

We trust that you will find the information presented within this document satisfactory. Should you have any questions or concerns, please do not hesitate to contact us at your convenience.

Sincerely,

Dillon Consulting Limited

Prepared by

Claire Smith, M.Sc.
Biologist

Reviewed by

Matt Gosse, B.Sc.
Associate, Project Manager

Susan Barfoot, P.Eng.
Associate

Attachments

A Site Photographs
B eDNA Lab Results

Attachment A

Site Photographs



Photo 1: Pond viewed from northwest.



Photo 2: Pond viewed from north.



Photo 3: Southern edge of pond, viewed from west.



Photo 4: Pond viewed from northeast.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 1,2,3,4



Photo 5: Northwestern shoreline of pond.



Photo 6: Northern shoreline of pond.



Photo 7: Southern shoreline from northeast.



Photo 8: Southern shoreline from southwest.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 5,6,7,8



Photo 9: Northeast shoreline.



Photo 10: Western shoreline.



Photo 11: Waterlily (Nuphar variegata).



Photo 12: Algae tufts.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 9,10,11,12



Photo 13: Southeastern shoreline.



Photo 14: Steep bank edges along southeastern shoreline.



Photo 15: Edge view of steep southeastern shoreline bank.



Photo 16: Close up of southeastern shoreline bank.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 13,14,15,16



Photo 17: Trench at northeast of pond, covered by boards.



Photo 18: Large boulders and berms at northeast of pond.



Photo 19: Concrete object near northern bank of pond.



Photo 20: Berms along eastern edge of pond.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 17,18,19,20



Photo 21: Another view of berms at eastern edge of pond.



Photo 22: Trench northeast of pond.



Photo 23: Trench northeast of pond with fence and sign.



Photo 24: Fence and sign northeast of pond.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 21,22,23,24



Photo 25: Biophysical habitat north of pond.



Photo 26: Biophysical habitat north of pond (2).



Photo 27: Biophysical habitat north of pond (3).



Photo 28: Biophysical habitat north of pond (4).

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 25,26,27,28



Photo 29: Biophysical habitat north of pond (5).



Photo 30: Biophysical habitat north of pond (6).



Photo 31: Biophysical habitat north of pond (7).



Photo 32: Biophysical habitat north of pond (8).

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 29,30,31,32



Photo 33: Red hose in ground north of pond.



Photo 34: Red hose in ground north of pond, close-up.



Photo 35: View of red hose in relation to pond.



Photo 36: Inside of red hose.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 33,34,35,36



Photo 37: Pond viewed from south.



Photo 38: Pond viewed from south (2).



Photo 39: Habitat south of pond.



Photo 40: Habitat south of pond, fallen tree.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 37,38,39,40



Photo 41: Habitat south of pond, continuation of grassy vegetat



Photo 42: Property marker south of pond, along eastern property edg



Photo 43: Flagging tape near property marker in Photo 42.



Photo 44: Vegetation and damp ground south of pond.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 41,42,43,44



Photo 45: Moose scat south of pond.



Photo 45: Potential moose tracks south of pond.



Photo 47: Potential moose tracks south of pond (2).



Photo 48: Potential moose tracks south of pond (3).

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 45,46,47,48



Photo 49: Biophysical habitat following potential southern outflow.



Photo 50: Biophysical habitat following potential southern outflow (2).



Photo 51: Biophysical habitat following potential southern outflow (3).



Photo 52: Wetland at potential southern outlet.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 49,50,51,52



Photo 53: Wetland at potential southern outlet (2).



Photo 54: Purple pitcher plant (Sarracenia purpurea).



Photo 55: Wetland at potential southern outlet (3).



Photo 56: Trail between nursery and pond.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 53,54,55,56



Photo 57: Nursery building roof viewed from trail.



Photo 58: Property marker northeast of pond.



Photo 59: Pipe entering pond from northwest.



Photo 60: Pipe entering pond from northwest (2).

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 57,58,59,60



Photo 61: Pipe entering pond from northwest (3).



Photo 62: Pipe entering pond from northwest (4).



Photo 63: Pipe entering pond from northwest (5).



Photo 64: Cap for water intake.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 61,62,63,64



Photo 65: Cap for water intake (2).



Photo 66: Cap for water intake (3).



Photo 67: Cap for water intake (4).



Photo 68: Cap for water intake (5).

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 65,66,67,68



Photo 69: Pump house.



Photo 70: Pump house, close-up.



Photo 71: Pump house, view of interior.



Photo 72: Black tubing section outside pump house.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 69,70,71,72



Photo 73: View of pump house from trail west of pond.



Photo 74: Property marker west of pond, northern edge of property.

DILLON	Site Photographs	Project No. 26-3638
07/07/2026	**101A Neary's Pond Rd Waterbody and Indeterminate Stream Assessment**	Photo No. 73,74

Appendix B

eDNA Lab Results

Attention: Matt Gosse
Dillon Consulting Ltd.
45 Hebron Way #202
St. John's, NL
A1A 0P9 Canada

Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
C.O.C. #: 20260618-1
Quote #: N/A
PO#: N/A

Report Date: 2026/06/24
Report #: DI20260624
Version: 1

ENVIRONMENTAL DNA - CERTIFICATE OF ANALYSIS

MXNS JOB #: E20260618-1
Received: 2026/06/18, 10:01 AM

Sample Type: Water
Samples Received: 3

Analyses (eDNA Isolation - Species)	Test Requested	Test Performed	Date eDNA Extracted	Date Conducted IntegritE-DNA™	Date Conducted Target Species	Laboratory Method	Analytical Method (qPCR Primer/Probe set)
eDNA Isolation and IntegritE-DNA™	3	3	2026/06/22	2026/06/22	N/A	GUE SOP-00056	ePlant5
American Eel (*Anguilla rostrata*)	3	3	N/A	N/A	2026/06/24	GUE SOP-00056	eANRO1
Brook trout (*Salvelinus fontinalis*)	3	3	N/A	N/A	2026/06/24	GUE SOP-00056	eSAFO6

Remarks:

The laboratory is accredited by Standards Council of Canada (SCC) to ISO/IEC 17025 for eDNA testing.

All work recorded herein has been done in accordance with procedures and practices ordinarily exercised by industry professionals using accepted testing methodologies, quality assurance and quality control procedures (except where otherwise agreed by the client and Merieux NutriSciences in writing). All data has met quality control and method performance criteria unless otherwise noted.

Merieux NutriSciences (MXNS) Laboratories' liability is limited to the actual cost of the requested analyses, unless otherwise agreed in writing. There is no other warranty expressed or implied. Merieux NutriSciences has been retained to provide analysis of samples provided by the Client using the testing methodology referenced in this report. Interpretation and use of test results are the sole responsibility of the Client and are not within the scope of services provided by Merieux NutriSciences unless otherwise agreed in writing. Merieux NutriSciences is not responsible for the accuracy or any data impacts that result from the information provided by the customer or their agent.

Results relate to supplied samples tested. This Certificate should not be reproduced except in full, without the written approval of the laboratory.

eDNA tests are used to confirm presence of eDNA in samples for the targeted species / species groups.

Collected eDNA samples will contain eDNA at various stages of degradation, being subject to environmental forces that breakdown DNA, including microbial activity, ultraviolet radiation, heat, hydrolysis, and enzymatic activity. eDNA is first evaluated for eDNA quality and presence of qPCR assay inhibitors using the IntegritE-DNA™ assay before testing for target species or genera to confirm that the eDNA is of sufficient quality for testing and to identify and address qPCR inhibition (if present) to avoid false negatives.

SAMPLE RETENTION: Samples and DNA extracts generated from the samples will be retained by Merieux NutriSciences for a period of 90 days after which time they will be discarded unless prearrangement has been made by client with Merieux NutriSciences for longer storage.

Attention: Matt Gosse
Dillon Consulting Ltd.
45 Hebron Way #202
St. John's, NL
A1A 0P9 Canada

Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
C.O.C. #: 20260618-1
Quote #: N/A
PO#: N/A

Report Date: 2026/06/24
Report #: DI20260624
Version: 1

ENVIRONMENTAL DNA - CERTIFICATE OF ANALYSIS

MXNS JOB #: E20260618-1
Received: 2026/06/18, 10:01 AM

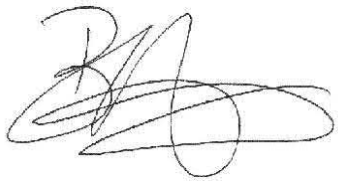
Methodology for Sample Analysis

Samples were logged in upon receipt. Water sample filtration and filter preservation were conducted at Merieux NutriSciences, Guelph location. The preserved samples were stored in freezer until processing in the laboratory. Sample analysis was completed within 10 business days (as indicated by the client on the COC) following receipt of samples by the testing laboratory.

eDNA isolation is completed using the DNeasy Blood & Tissue Kit™ (QIAGEN). A negative control is included as a blank filter sample with each batch of eDNA isolation to monitor for potential laboratory contamination during the eDNA isolation process.

Following eDNA isolation (150µL) from a quarter of filter, the IntegritE-DNA™ assay[1] is used to avoid the potential of a false negative (Type II error) during target species or genera testing. The IntegritE-DNA™ assay evaluates the integrity of eDNA for suitability for qPCR and for presence of qPCR inhibitors which may reduce the effectiveness of the qPCR assay for target species or genera. This assay evaluates the quality of eDNA to assess whether it is amplifiable using a qPCR assay that targets the chloroplast genome derived from plants/algae that are ubiquitously found in fresh water systems. Four technical replicates per eDNA sample, four technical replicates of negative control (Ultrapure water), and two technical replicates of positive control are used for the IntegritE-DNA™ assay. The cut-off Ct (qPCR cycle threshold) value for the IntegritE-DNA™ assay is 27 due to inhibition. If the IntegritE-DNA™ assay produces a positive detection frequency of ≥ 2 of the 4 technical replicates, this indicates that the eDNA for the target taxa is likely to be of sufficient quality to be detected (if present) with the target assay. If the IntegritE-DNA™ assay produces a positive detection frequency < 2 of the 4 technical replicates (eDNA is degraded or qPCR inhibitors are present), then sample cleanup is completed using the OneStep PCR Inhibitor Removal Kit™ (ZYMO Research) to remove potential qPCR assay inhibitors from the isolated eDNA. Subsequent to inhibitor removal, the IntegritE-DNA™ assay is repeated to re-assess whether the eDNA is of sufficient quality for qPCR. If a sample fails at the IntegritE-DNA™ assay (Ct value over 30) for the second time the client will be informed that the quality of the sample is insufficient for the qPCR assay. eDNA indicator (IntegritE-DNA™) in the sample suggests that degradation has taken place and therefore the target species assay may be ineffective. Once a sample passes the IntegritE-DNA™ assay, then the target species or genera assay is performed. Eight technical replicates per eDNA sample, eight technical replicates of the negative control (Ultrapure water), and two technical replicates of positive control (total DNA or synthetic DNA) are used for the target species or genera assay to assess the detection or non-detection of DNA of the target species or genera. The cut-off Ct value for target species assay is 50.

[1] Hobbs J, Round JM, Allison MJ, Helbing CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849.



BECKY HENDERSON
Customer Care Representative Premier, Merieux NutriSciences, DNA Services
Email: becky.a.henderson@mxns.com
Phone #: (519) 836 2400 Ext. 7067714

Please direct all questions regarding this Certificate of Analysis to your Customer Service Representative above.

For Service Group specific validation please refer to the Validation Signature Page.

Total Cover Pages: 2

Mérieux NutriSciences

MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

RESULTS - American Eel (*Anguilla rostrata*)

Client Sample ID	MXNS Case ID	Sampling Date	IntegritE-DNA™ Positive detection (Ct≤27)[1]	QC Batch	Clean-up required	IntegritE-DNA™ Positive detection (Ct≤30)[1] after clean-up	QC Batch	Analytical Method (qPCR Primer/Probe set)	Target Species eDNA Positive detection (Ct≤50)[2]	QC Batch
26-EDNA01	DI20260122	2026/06/17	4/4	260622Q1	No	N/A	N/A	eANRO1[4]	0/8	260624Q2
26-EDNA02	DI20260123	2026/06/17	4/4	260622Q1	No	N/A	N/A	eANRO1	0/8	260624Q2
26-EDNA03	DI20260124	2026/06/17	4/4	260622Q1	No	N/A	N/A	eANRO1	0/8	260624Q2
Filtration Blank Before	FBB260618	2026/06/18	0/4[3]	260622Q1	No[3]	N/A	N/A	eANRO1	N/A	N/A
Filtration Blank After	FBA260618	2026/06/18	0/4[3]	260622Q1	No[3]	N/A	N/A	eANRO1	N/A	N/A

[1] **IntegritE-DNA™ Assay:** Four technical replicates were assayed for each eDNA sample. **Field samples:** The cut-off Ct value for IntegritE-DNA™ assay is 27 based on previous observation and due to presence of inhibitors. **Post clean-up and field blank samples:** The cut-off Ct value for IntegritE-DNA™ assay is 30. Results are reported as the number of positive detections (n) out of a total of 4 technical replicates, n/4.

[2] **Target Species Assay:** Eight technical replicates were assayed per eDNA sample. The cut-off Ct value for target species assay was 50. Results are reported as the number of positive detections (n) out of a total of 8 technical replicates, n/8.

[3] The IntegritE-DNA assay failed for Filtration Blank samples as expected. Clean-up is not required.

[4] eANRO1: qPCR primer/probe assay to assess the presence of American Eel (*Anguilla rostrata*) eDNA.

RESULTS - Brook trout (*Salvelinus fontinalis*)

Client Sample ID	MXNS Case ID	Sampling Date	IntegritE-DNA™ Positive detection (Ct≤27)[1]	QC Batch	Clean-up required	IntegritE-DNA™ Positive detection (Ct≤30)[1] after clean-up	QC Batch	Analytical Method (qPCR Primer/Probe set)	Target Species eDNA Positive detection (Ct≤50)[2]	QC Batch
26-EDNA01	DI20260122	2026/06/17	4/4	260622Q1	No	N/A	N/A	eSAFO6[4]	0/8	260624Q4
26-EDNA02	DI20260123	2026/06/17	4/4	260622Q1	No	N/A	N/A	eSAFO6	0/8	260624Q4
26-EDNA03	DI20260124	2026/06/17	4/4	260622Q1	No	N/A	N/A	eSAFO6	0/8	260624Q4
Filtration Blank Before	FBB260618	2026/06/18	0/4[3]	260622Q1	No[3]	N/A	N/A	eSAFO6	N/A	N/A
Filtration Blank After	FBA260618	2026/06/18	0/4[3]	260622Q1	No[3]	N/A	N/A	eSAFO6	N/A	N/A

[1] **IntegritE-DNA™ Assay:** Four technical replicates were assayed for each eDNA sample. **Field samples:** The cut-off Ct value for IntegritE-DNA™ assay is 27 based on previous observation and due to presence of inhibitors. **Post clean-up and field blank samples:** The cut-off Ct value for IntegritE-DNA™ assay is 30. Results are reported as the number of positive detections (n) out of a total of 4 technical replicates, n/4.

[2] **Target Species Assay:** Eight technical replicates were assayed per eDNA sample. The cut-off Ct value for target species assay was 50. Results are reported as the number of positive detections (n) out of a total of 8 technical replicates, n/8.

[3] The IntegritE-DNA assay failed for Filtration Blank samples as expected. Clean-up is not required.

[4] eSAFO6: qPCR primer/probe assay to assess the presence of Brook trout (*Salvelinus fontinalis*) eDNA.

MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

GENERAL COMMENTS

eDNA is extracted (150 µL) from a quarter of filter, and 2 µL is used as a template for each technical replicate.

Two blank samples (Ultra-Pure water) were filtered before and after filtration (each batch) as a control for contamination. All filtration blanks did not detect any DNA by IntegritE-DNA assay as expected.

Results relate only to the items tested.

QUALITY ASSURANCE REPORT

			eDNA Isolation Negative Control[1]		qPCR Positive Controls[2]		qPCR Negative Controls[3]	
QC Batch	**Parameter**	**Date**	**Detection at: Ct 27 (IntegritE-DNA™) Ct 50 (other assays)**	**Pass / Fail**	**Detection at: Ct 27 (IntegritE-DNA™) Ct 50 (other assays)**	**Pass / Fail**	**Detection at: Ct 27 (IntegritE-DNA™) Ct 50 (other assays)**	**Pass / Fail**
260622Q1	IntegritE-DNA	2026/06/22	0 of 4 technical replicates	Pass	2 of 2 technical replicates	Pass	0 of 4 technical replicates	Pass
260624Q2	eANRO1	2026/06/24	eDNA Isolation Negative Control is assessed using IntegritE-DNA only once for each extraction batch.	N/A	2 of 2 technical replicates	Pass	0 of 8 technical replicates	Pass
260624Q4	eSAFO6	2026/06/24			2 of 2 technical replicates	Pass	0 of 8 technical replicates	Pass

[1]**eDNA Isolation Negative Control:** Blank filters were included for each batch of eDNA extraction to monitor for laboratory contamination during eDNA isolation. eDNA Isolation Negative Control is assessed using IntegritE-DNA™ only. QC results show no eDNA was isolated from the negative control, therefore there was no indication of sample contamination during handling. Acceptance criteria: 0 of 4 technical replicates

[2]**qPCR Positive Controls:** Two technical replicates of isolated eDNA from freshwater sample were used as positive controls for IntegritE-DNA™. Two technical replicates of total DNA or synthetic DNA from the target species were used as positive controls for eDNA assays. Results show that 100% of the technical replicates amplified the positive control eDNA as expected, therefore an observation of negative result in eDNA samples is not related to the qPCR performance. Acceptance criteria: 2 of 2 technical replicates

[3]**qPCR Negative Controls (Ultrapure water):** Four technical replicates for IntegritE-DNA™ and eight technical replicates for target species or genera were used to monitor for laboratory contamination. Results show that 0% of the technical replicates in the negative controls had amplified eDNA, indicating no contamination was detected. Acceptance criteria: 0 of 4 technical replicates for IntegritE-DNA™, and 0 of 8 technical replicates for other assays.

LABORATORY RESULTS VALIDATION SIGNATURE PAGE

The analytical data and all QC contained in this report were reviewed and validated by the following individual(s).

Reporter: ALI MIRABZADEH, M.Sc.
Operations Supervisor, Merieux NutriSciences, DNA Services

Mérieux NutriSciences

MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

Brook trout (*Salvelinus fontinalis*) eDNA Assay Validation Information

eDNA assay Validation

All eDNA assays are validated through a rigorous multi-step evaluation protocol at University of Victoria that includes tests of DNA target specificity and amplification sensitivity. All eDNA tests available at Merieux NutriSciences have been internally verified using genomic DNA and/or field positive samples.

General eDNA Assay Information

Target Species: Brook trout (*Salvelinus fontinalis*)
eDNA qPCR Tool: eSAFO6
Gene Target: MT-CO3
Species Code: te-SAFO
eDNA qPCR Format: TaqMan
Published in: N/A

eDNA Assay Sensitivity Test using gBlocks™ synthetic DNA (Merieux NutriSciences)

LOD **0.54** 95% CI **0.35-1.2** Copies/Rxn LOQ **2.05** 95% CI **1.33-4.56** Copies/Rxn
LOB **0** hits/8 LOQcontinuous **4** Copies/Rxn Enzyme: Immolase

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code[4]. When the LOQ < LOD, use the LOD for the LOQ.

eDNA Assay Specificity Test Information (University of Victoria, Dr. Caren Helbing Laboratory)

Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Species	Common Name (Species)	Detection	# Voucher Specimens	Sample Sources/Locations
ma-CALUfa	Dog (*Canis lupus familiaris*)	No	1	British Columbia
ma-HOSA	Human (*Homo sapiens*)	No	1	Netherlands
te-COAR	Cisco/Tullibee (*Coregonus artedi*)	No	2	Alberta
te-COCL	Lake whitefish (*Coregonus clupeaformis*)	No	1	Alberta
te-ESLU	Northern Pike (*Esox lucius*)	No	1	British Columbia
te-MIDO	Smallmouth Bass (*Micropterus dolomieu*)	No	2	British Columbia
te-MISA	Largemouth Bass (*Micropterus salmoides*)	No	2	British Columbia
te-ONCLle	Westslope cutthroat trout (*Oncorhynchus clarki lewisi*)	No	2	Alberta
te-ONGO	Pink Salmon (*Oncorhynchus gorbuscha*)	No	2	British Columbia
te-ONKE	Chum Salmon (*Oncorhynchus keta*)	No	2	British Columbia
te-ONKI	Coho Salmon (*Oncorhynchus kisutch*)	No	2	British Columbia
te-ONMY	Rainbow Trout (*Oncorhynchus mykiss*)	No	2	Alberta
te-ONTS	Chinook Salmon (*Oncorhynchus tschawytscha*)	No	1	British Columbia
te-PRCY	Round Whitefish (*Prosopium cylindraceum*)	No	1	Yukon
te-SACO	Bull Trout (*Salvelinus confluentus*)	No	2	Alberta
te-SAFO	Brook trout (*Salvelinus fontinalis*)	Yes	5	Alberta
te-SAMA	Dolly Varden (*Salvelinus malma*)	No	2	British Columbia
te-SANA	Lake trout (*Salvelinus namaycush*)	No	2	Alberta
te-SASA	Atlantic Salmon (*Salmo salar*)	No	1	Nova Scotia
te-THAR	Arctic Grayling (*Thymallus arcticus*)	No	2	Alberta

Field Sample Validation

Sample Type	Known presence	# Samples	Detected	Location
N/A	N/A	N/A	N/A	N/A

Abbreviations

95% CI	95% Confidence interval	LOQ	Limit of quantification
eDNA	Environmental DNA	MT-CO3	Mitochondrial cytochrome oxidase subunit 3 gene
gDNA	Total genomic DNA extracted from voucher specimen	NTC	qPCR no template control
LOB	Limit of blank	qPCR	Quantitative real-time polymerase chain reaction
LOD	Limit of detection	SE	Standard error

Mérieux NutriSciences

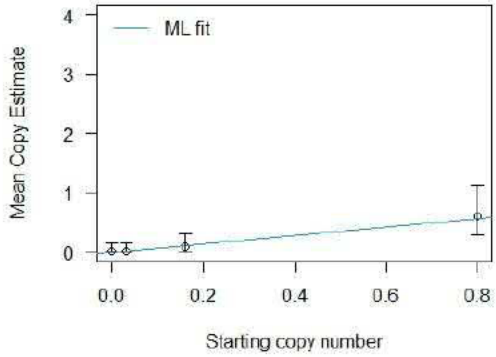
MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

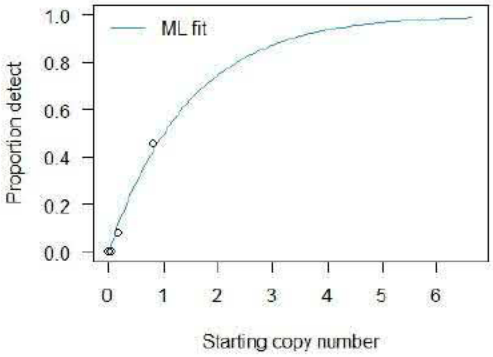
Brook trout (*Salvelinus fontinalis*) eDNA Assay Validation Information

eDNA Assay Sensitivity Test Details using gBlocks™ synthetic DNA (Merieux NutriSciences)

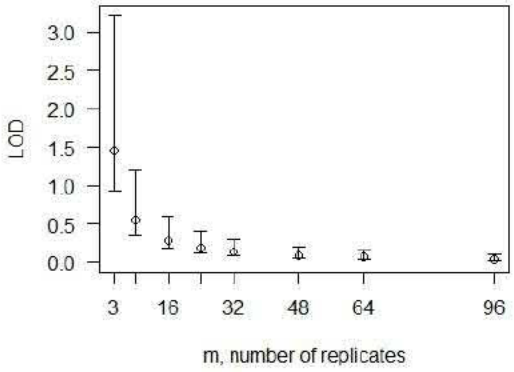
eSAFO6



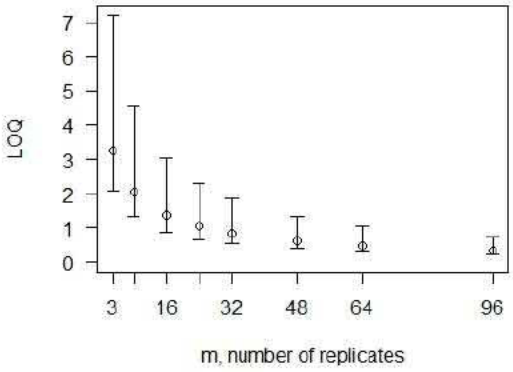
No intercept eSAFO6



Limits detect - no intercept eSAFO6



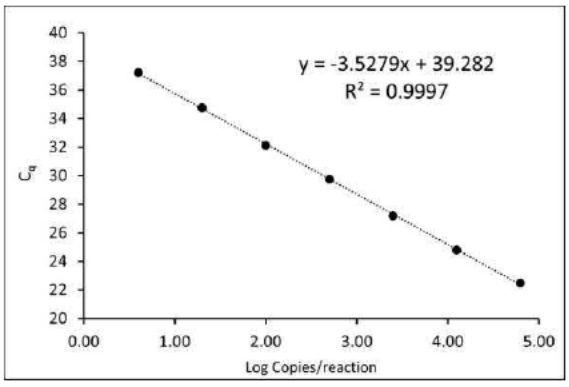
Limits quant - no intercept eSAFO6



Binomial-Poisson model: **No intercept**

Determined using eLowQuant R code[4].

Based on a 2 µL DNA input in a total 15 µL reaction



Efficiency **92%**

Mérieux NutriSciences

MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

Brook trout (*Salvelinus fontinalis*) eDNA Assay Validation Information

eDNA Assay Sensitivity Test Details using gBlocks™ synthetic DNA (Merieux NutriSciences)

# Detects	Number of Technical Replicates							
	n=8		n=16		n=24		n=32	
	# Copies	SE	# Copies	SE	# Copies	SE	# Copies	SE
0	0.000	N/A	0.000	N/A	0.000	N/A	0.000	N/A
1	0.194	0.201	0.094	0.097	0.062	0.064	0.046	0.048
2	0.417	0.318	0.194	0.147	0.126	0.096	0.094	0.071
3	0.681	0.441	0.301	0.193	0.194	0.124	0.143	0.092
4	1.005	0.585	0.417	0.240	0.264	0.152	0.194	0.111
5	1.422	0.772	0.543	0.288	0.339	0.179	0.246	0.130
6	2.011	1.053	0.681	0.340	0.417	0.207	0.301	0.149
7	3.014	1.597	0.834	0.396	0.500	0.236	0.358	0.169
8			1.005	0.459	0.588	0.266	0.417	0.189
9			1.198	0.531	0.681	0.298	0.479	0.209
10			1.422	0.615	0.781	0.332	0.543	0.230
11			1.686	0.716	0.889	0.369	0.611	0.252
12			2.009	0.843	1.005	0.409	0.681	0.275
13			2.426	1.016	1.131	0.452	0.756	0.300
14			3.014	1.278	1.269	0.499	0.834	0.325
15			4.019	1.800	1.422	0.552	0.917	0.352
16					1.592	0.612	1.005	0.381
17					1.786	0.681	1.098	0.411
18					2.009	0.762	1.198	0.444
19					2.274	0.860	1.306	0.480
20					2.597	0.984	1.422	0.518
21					3.014	1.152	1.548	0.560
22					3.602	1.408	1.686	0.607
23					4.606	1.919	1.839	0.658
24							2.010	0.717
25							2.203	0.785
26							2.426	0.864
27							2.691	0.961
28							3.014	1.083
29							3.431	1.249
30							4.016	1.500
31							5.023	2.005

Binomial-Poisson model, determined using eLowQuant R code[4].

LOD and LOQ with confidence intervals, by number of replicates, no intercept model

	3	8	16	24	32	48	64	96
LOD	1.45	0.54	0.27	0.18	0.14	0.09	0.07	0.05
LOD_lower	0.93	0.35	0.18	0.12	0.09	0.06	0.04	0.03
LOD_upper	3.21	1.20	0.60	0.40	0.30	0.20	0.15	0.10
LOQ	3.24	2.05	1.36	1.03	0.84	0.61	0.48	0.34
LOQ_lower	2.09	1.33	0.88	0.67	0.54	0.39	0.31	0.22
LOQ_upper	7.19	4.56	3.03	2.30	1.86	1.35	1.06	0.74

References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus* , in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. https://doi.org/10.1002/edn3.82
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei* , in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. https://doi.org/10.1371/journal.pone.0213849
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 00: 1-12. doi: 10.1002/edn3.220

Mérieux NutriSciences

MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

American Eel (*Anguilla rostrata*) eDNA Assay Validation Information

eDNA assay Validation

All eDNA assays are validated through a rigorous multi-step evaluation protocol at University of Victoria that includes tests of DNA target specificity and amplification sensitivity. All eDNA tests available at Merieux NutriSciences have been internally verified using genomic DNA and/or field positive samples.

General eDNA Assay Information

Target Species: American Eel (*Anguilla rostrata*)
Species Code: te-ANRO
eDNA qPCR Tool: eANRO1
eDNA qPCR Format: TaqMan
Gene Target: MT-CYB
Published in: N/A

eDNA Assay Sensitivity Test using gBlocks™ synthetic DNA (University of Victoria, Dr. Caren Helbing Laboratory)

LOD **0.4** 95% CI **0.3-0.7** Copies/Rxn LOQ **1.5** 95% CI **1.1-2.6** Copies/Rxn
LOB **0** hits/8 LOQcontinuous **20** Copies/Rxn Enzyme: Immolase

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code[4].

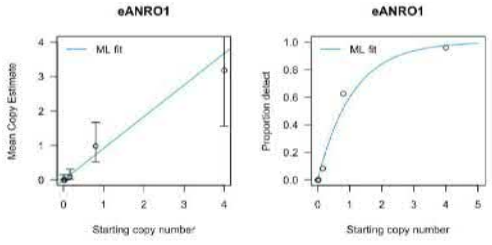
When the LOQ < LOD, use the LOD for the LOQ.

eDNA Assay Specificity Test Information (University of Victoria, Dr. Caren Helbing Laboratory)

Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Species	Common Name (Species)	Detection	Specimens	Sample Sources/Locations
ma-HOSA	Human (*Homo sapiens*)	No	1	Netherlands
te-ANRO	American Eel (*Anguilla rostrata*)	Yes	8	Prince Edward Island
te-COCO	Slimy Sculpin (*Cottus cognatus*)	No	1	Yukon
te-ESLU	Northern Pike (*Esox lucius*)	No	1	British Columbia
te-MIDO	Smallmouth Bass (*Micropterus dolomieu*)	No	1	British Columbia
te-MISA	Largemouth Bass (*Micropterus salmoides*)	No	1	British Columbia
te-ONCL	Cutthroat Trout (*Oncorhynchus clarkii*)	No	1	British Columbia
te-ONGO	Pink Salmon (*Oncorhynchus gorbuscha*)	No	1	British Columbia
te-ONKE	Chum Salmon (*Oncorhynchus keta*)	No	1	British Columbia
te-ONKI	Coho Salmon (*Oncorhynchus kisutch*)	No	1	British Columbia
te-ONMY	Rainbow Trout (*Oncorhynchus mykiss*)	No	1	British Columbia
te-ONNE	Sockeye Salmon (*Oncorhynchus nerka*)	No	1	British Columbia
te-ONTS	Chinook Salmon (*Oncorhynchus tschawytscha*)	No	1	British Columbia
te-PRCY	Round Whitefish (*Prosopium cylindraceum*)	No	1	Yukon
te-SACO	Bull Trout (*Salvelinus confluentus*)	No	1	Alberta
te-SAMA	Dolly Varden (*Salvelinus malma*)	No	1	British Columbia
te-SASA	Atlantic Salmon (*Salmo salar*)	No	1	Nova Scotia
te-THAR	Arctic Grayling (*Thymallus arcticus*)	No	1	Alberta
te-THPA	Eulachon (*Thaleichthys pacificus*)	No	1	British Columbia

eDNA Assay Sensitivity Test using gBlocks™ synthetic DNA (University of Victoria, Dr. Caren Helbing Laboratory)



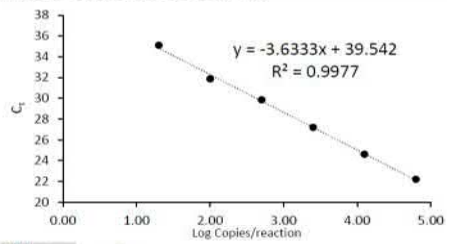
From 8 Technical Replicates

# Detects	# Copies	SE
0	0	0
1	0.15	0.14
2	0.31	0.23
3	0.51	0.32
4	0.76	0.41
5	1.07	0.54
6	1.51	0.73
7	2.27	1.12

Determined using eLowQuant R code[4].

Binomial-Poisson model: No intercept
Determined using eLowQuant R code[4].
Based on a 2 µL DNA input in a total 15 µL reaction

Applied to reactions with 100% positive hits



Efficiency 88%

American Eel (*Anguilla rostrata*) eDNA Assay Validation Information

Field Sample Validation

Sample Type	Known presence	# Samples	Detected	Location
Water	Yes	30	Y	Prince Edward Island

Abbreviations

95% CI	95% Confidence interval	LOQ	Limit of quantification
eDNA	Environmental DNA	MT-CYB	Mitochondrial cytochrome B gene
gDNA	Total genomic DNA extracted from voucher specimen	NTC	qPCR no template control
LOB	Limit of blank	qPCR	Quantitative real-time polymerase chain reaction
LOD	Limit of detection	SE	Standard error

References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus* , in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. https://doi.org/10.1002/edn3.82
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei* , in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. https://doi.org/10.1371/journal.pone.0213849
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 00: 1-12. doi: 10.1002/edn3.220

SUPPLEMENTARY INFORMATION

eDNA Isolation Method:	☑ DNeasy Blood & Tissue Kit™ (QIAGEN) ☐ Modified* DNeasy Blood & Tissue Kit™ (QIAGEN)
eDNA Isolation Negative Control:	☑ Cellulose Nitrate Membrane Filter (0.45 µm, 47 mm)
qPCR Negative Control (NTC):	☑ Ultra-pure water is used as a template.
qPCR Positive Control:	☑ Synthetic DNA (gBlocks) and/or Genomic DNA (gDNA)
qPCR Enzyme:	☑ IMMOLASE™ DNA Polymerase ☐ QIAcuity® Probe PCR Kit
Clean-up Kit:	☐ OneStep PCR Inhibitor Removal Kit™ (ZYMO Research)
Real-Time Thermal Cycler:	☑ Bio-Rad, CFX96 Touch Real-Time PCR ☐ ABI, 7500 Real-Time PCR System (v1.2 build 31)
Data Analysis Software:	☑ Bio-Rad, CFX Maestro Version# 2.3 ☐ ABI, SDS Software, v1.2 build 29

* The modified method of extraction is similar to DNeasy Blood & Tissue Kit™ (QIAGEN) method with one exception. The ATL buffer (lysis buffer) has been replaced by InhibitEx Buffer from the QIAamp Fast DNA Stool Mini Kit (QIAGEN). InhibitEx Buffer efficiently depletes PCR inhibitors from sample during the extraction, enabling reliable detection of DNA.

Mérieux NutriSciences

MXNS JOB #: E20260618-1
Report Date: 2026/06/24
Report #: DI20260624
Version #: 1

Client Name: Dillon Consulting Ltd.
Client Project #: 26-3686
Site Location: Portugal Cove-St Philips
Sampler Initials: N/A

Mérieux NutriSciences

ENVIRONMENTAL DNA (eDNA) CHAIN OF CUSTODY RECORD

«An incomplete or incorrect form may lead to delays in testing»

Send the samples to:
Mérieux NutriSciences, DNA Services
335 Laird Rd #2
Guelph, ON, Canada N1G 4P7
eDNA-na@mxns.com

For Lab Use Only
COC#: 20260618-1

Page: _____ of _____

1 Invoice Information (Required; Accounts Payable)		2 Report Information		3 Project Information (where applicable)	
Company Name:	Dillon Consulting	Company Name:	Dillon Consulting	Quotation #:	
Contact Name:	Jessica Brinston	Contact Name:	Matt Gosse	P.O. #:	
Address:	45 Hebron Way	Address:	45 Hebron Way	Project #:	26-3686
	St. John's, NL		St. John's, NL	Site Location:	Portugal Cove-St. Philips
Phone:	709-727-3228	Phone:	709-727-3228	Note:	
Email:	jbrinston@dillon.ca	Email:	mgosse@dillon.ca		

4 Turnaround Time (TAT) (Required)

Regular TAT		Rush TAT (Surcharges will be applied) EMAIL eDNAna@mxns.com FOR RUSH PROJECTS	
☑	10 business days based upon a maximum of 50 samples received per batch. For every additional increment of 50 samples, please allow an extra 5 business days.	☐	5 business days based upon a maximum of 50 samples received per batch. For every additional increment of 50 samples, please allow an extra 3 business days.

eDNA ANALYSIS

5 IMPORTANT INFORMATION

- Water samples should be kept cool and filtered within 24 hours of collection to ensure optimal results.
- For eDNA testing, it is recommended to use a Cellulose Nitrate (CN) filter.
- After filtration, preserve the filter immediately by storing it with self-indicating silica beads.

6 CLIENT'S SPECIAL INSTRUCTIONS

Please filter in lab

7 Number	7 Sample Identification	8 Date Sampled (YYYY/MM/DD)	9 Date Filtered and Preserved (YYYY/MM/DD)	10 Filter Material	11 Filter Size (Diameter)	12 Filter Pore Size (µm)	13 Preservation Method	14 Assays Requested*	15 Comments
1	26-EDNA01	26 06 17						Brook trout & American Eel	
2	26-EDNA02	26 06 17						Brook trout & American Eel	
3	26-EDNA03	26 06 17						Brook trout & American Eel	
4									
5									
6									
7									
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9									
10									
11									
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18									
19									
20									

16 RELINQUISHED BY: (Signature/Print)	17 DATE: (YYYY/MM/DD)	18 TIME: (HH:MM)	RECEIVED BY: (Signature/Print)	DATE: (YYYY/MM/DD)	TIME: (HH:MM)
Matt Gosse	2026/06/17	13:00	BHAM	For Lab Use Only JUN 18 2026	10:21

* Email ... to receive the list of available eDNA assays at Mérieux NutriSciences.